





Tokyo University of Pharmacy and Life Sciences.

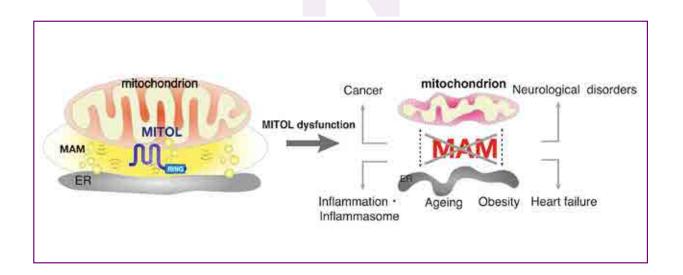
I am the representative for the research program entitled "Roles of membrane contact sites in organelle dynamics and diseases" adopted as a MEXT-Supported Program for the Strategic Research Foundation at Private Universities in 2014.

Cell is the structural and functional unit of life. In the cell, there are several kind of organelles surrounded with membrane and they closely communicate with each other. It is necessary for the efficient signal transmission between different organelles to collect information at proximal position without dispersion. Thus, regulation of membrane contact sites in organelle dynamics seems to be important for maintenance of cell function. In particular, biochemical and recent electron microscopy (EM) studies confirmed the presence of specific regions of close apposition between mitochondria and ER, termed mitochondria-associated membranes (MAM). MAM plays a role in many cellular functions, including calcium homeostasis, phospholipid metabolism, formations of autophagosome and inflammasome, apoptosis and cell growth signaling. Furthermore, it has been reported that MAM dysfunction is involved in various diseases such as Alzheimer disease, cardiac failure, cancer, and anti-virus immune response. Thus, it is emerging project to research MAM function for understanding of disease pathology and development of new drug against these diseases.



Roles of Membrane Contact Sites in Organelle Dynamics and Diseases

We can challenge for elucidation of MAM function under the circumstances, because excellent scientists with a proven track record for MAM research gather in School of life Sciences of Tokyo university of Pharmacy and life Sciences. In cooperation with many experts for various fields in the inside and outside of university, we try to clarify the roles of MAM in organelle dynamics and diseases. This project may contribute greatly to understanding of pathology and development of new treatments for MAM-associated diseases. In addition, we aim for cultivation of young scientists who are responsible for next generation.





CONTENTS

P.1~2

Greeting from the Organizer

P.3

Contents

P.4~9

Group for Basic Molecular Mechanism Activity Report



Shigeru Yanagi, M.D., Ph.D.
Professor
Laboratory of Molecular Biochemistry.
School of Life Sciences,
Tokyo University of
Pharmacy and Life Sciences.





Shun Nagashima, Ph.D. Assistant Professor Laboratory of Molecular Biochemistry, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.



Mitsuo Tagaya, Ph.D.
Professor
Laboratory of Molecular Cell Biology,
School of Life Sciences,
Tokyo University of
Pharmacy and Life Sciences.



Katsuko Tani, Ph.D.
Professor
Laboratory of Cell Signaling,
School of Life Sciences,
Tokyo University of
Pharmacy and Life Sciences.
P.6



Kohei Arasaki, Ph.D. Assistant Professor Laboratory of Molecular Cell Biology, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.



Takashi Baba, Ph.D.
Assistant Professor
Laboratory of Cell Signaling,
School of Life Sciences,
Tokyo University of
Pharmacy and Life Sciences.
P.9



Group for molecular mechanism of diseases Activity Report



Kiyoko Fukami, Ph.D.
Professor
Laboratory of Genome and Biosignals,
Tokyo University of
Pharmacy and Life Science.

P.10



Tetsuro Watabe, Ph.D.
Professor
Laboratory of Oncology,
School of Life Sciences,
Tokyo University of
Pharmacy and Life Science.
P.12



Masato Tanaka, M.D., Ph.

Professor
Laboratory of Immune Regulation,
School of Life Sciences,
Tokyo University of
Pharmacy and Life Sciences.

P.11



Yoshitami Hashimoto, Ph.D. Assistant Professor Laboratory of Cellular Regulation, Tokyo University of Pharmacy and Life Sciences.

P.13



Private Universities (2014-2018)

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases

Elucidation of the roles of MITOL in the formation of mitochondria-associated membrane (MAM) and relationship between MAM disruption and diseases



Shigeru Yanagi, M.D., Ph.D.

Professor

Laboratory of Molecular Biochemistry, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.

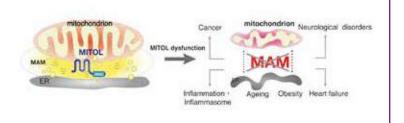
Research summary

The mitochondrion plays an important role in exchanging Ca2+ or metabolizing lipids by positioning at sites situated close to the endoplasmic reticulum (ER), which is called MAM. Mitofusin2 (Mfn2) is a mitochondrial fusion factor which is localized in the endoplasmic reticulum as well as in mitochondria and play a role in the attachment of these organelles to each other. Although a previous study indicates that Mfn2 is required for MAM formation, the mechanisms regulating this process remain unknown. Recently we have demonstrated that MITOL ubiquitinates Mfn2 and induces MAM formation through Mfn2 oligomerization. However, the regulatory mechanism for MITOL-induced MAM formation and the involvement of disrupted MAM in disease pathology are still obscure. In this project, we will explore the roles of MITOL in the MAM formation and MAM-associated diseases such as Alzheimer disease.

Figure

A model of the mechanism by which MITOL regulates MAM formation

MITOL ubiquitinates mitochondrial Mfn2 on its lysine residue in position 192 in a K63-dependent manner, thereby activating Mfn2. In turn, activated Mfn2 binds to Mfn2 localized in the ER, resulting in mitochondria-ER tethering by oligomerization of Mfn2.



- 1.Sugiura, A., Nagashima, S., Tokuyama, T., Amo, T., Matsuki, Y., Ishido, S., Kudo, Y., McBride, H.M., Fukuda, T., Matsushita, T., Inatome, R., and *Yanagi, S. (2013) MITOL regulates endoplasmic reticulum-mitochondria contacts via Mitofusin2. *Molecular Cell.* 51:1-15.
- 2. Yonashiro, R., Kimijima, Y., Shimura, T., Kawaguchi, K., Fukuda, T., Inatome, R., and *Yanagi, S. (2012) Mitochondrial ubiquitin ligase MITOL blocks S-nitrosylated MAP1B-light chain 1-mediated mitochondrial dysfunction and neuronal cell death. *Proceedings of the National Academy of Sciences U.S.A.* 109:2382-2387.
- 3.Sugiura, A., Yonashiro, R., Fukuda, T., Matsushita, N., Nagashima, S., Inatome, R., and *Yanagi, S. (2011) A mitochondrial ubiquitin ligase MITOL controls cell toxicity of polyglutamine-expanded protein. *Mitochondrion*. 11:139-146.
- 4. Yonashiro, R., Sugiura, A., Miyachi, M., Fukuda, T., Matsushita, N., Inatome, R., Ogata, Y., Suzuki, T., Dohmae, N., and *Yanagi, S. (2009) Mitochondrial ubiquitin ligase MITOL ubiquitinates mutant SOD1 and attenuates mutant SOD1-induced ROS generation. *Molecular Biology of the Cell*. 20:4524-4530
- 5. Yonashiro, R., Ishido, S., Kyo, S., Fukuda, T., Goto, E., Matsuki, Y., Ohmura-Hoshino, M., Sada, K., Hotta, H., Yamamura, H., Inatome, R., and *Yanagi, S. (2006) A novel mitochondrial ubiquitin ligase plays a critical role in mitochondrial dynamics. *EMBO Journal*. 25:3618-3626.



Diseases

Roles of Membrane Contact Sites in Organelle Dynamics and

Elucidation of the roles of syntaxin 17 localized in the mitochondria-associated membrane (MAM) and its participation in MAM-associated diseases



Mitsuo Tagaya, Ph.D.

Professor

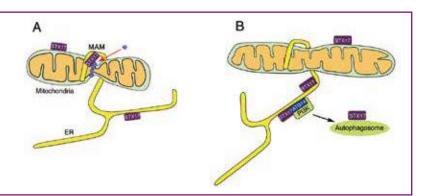
Laboratory of Molecular Cell Biology, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.

Research summary

The endoplasmic reticulum (ER) contains various subdomains that are in contact with other organelles. The ER subdomain facing mitochondria is called the mitochondria-associated membrane (MAM). The MAM regulates mitochondrial activity through Ca²+ and synthesizes lipids in cooperation with mitochondria. Accumulating data have disclosed that the ER-mitochondria interface is the site for various important cell functions, beyond Ca²+ homeostasis and lipid synthesis. Moreover, the close relationship between this site and neurodegenerative diseases has been pointed out. In this project, we will explore the roles of syntaxin 17 (STX17) in the ER-mitochondria interface and MAM-associated diseases.

Figure

Different roles of STX17 in response to cellular physiology. (A) In fed cells, STX17 promotes mitochondrial fission by regulating Drp1 (represented by D in blue circle) localization/activity. (B) In starved cells, STX17 switches its binding from Drp1 to the PI3K subunit ATG14, leading to mitochondrial elongation and autophagosome formation. This elongation allows mitochondria to escape from autophagic degradation.



- 1.Arasaki K, Shimizu H, Mogari H, Nishida N, Hirota N, Furuno A, Kudo Y, Baba M, Baba N, Cheng J, Fujimoto T, Ishihara N, Ortiz-Sandoval C, Barlow LD, Raturi A, Dohmae N, Wakana Y, Inoue H, Tani K, Dacks JB, Simmen T, *Tagaya M. (2015) Developmental Cell. 32:304-317.
- 2.Noda C, Kimura H, Arasaki K, Matsushita M, Yamamoto A, Wakana Y, Inoue H, *Tagaya, M. (2014) Valosin-containing protein-interacting membrane protein (VIMP) links the endoplasmic reticulum with microtubules in concert with cytoskeleton-linking membrane protein (CLIMP)-63. *Journal of Biological Chemistry*. 289:24304-24313.
- 3.* Tagaya M. Arakaki K, Inoue H, Kimura H. (2014) Moonlighting functions of the NRZ (mammalian Dsl1) complex. Frontiers in Cell and Developmental Biology. 2:25.
- 4.He S, Ni D, Ma B, Lee JH, Zhang T, Ghozalli I, Pirooz SD, Zhao Z, Bharatham N, Li B, Oh S, Lee WH, Takahashi Y, Wang HG, Minassian A, Feng P, Deretic V, Pepperkok R, <u>Tagaya M</u>, Yoon HS, *Liang C. (2013) PI(3)P-bound UVRAG coordinates Golgi-ER retrograde and Atg9 transport by differential interactions with the ER tether and the Beclin1 complex. *Nature Cell Biology* 15:1206-1219.
- 5.Arasaki K, Takagi D, Furuno A, Sohda M, Misumi Y, Wakana Y, Inoue H, *Tagaya M. (2013) A new role for RINT-1 in SNARE complex assembly at the trans-Golgi network in coordination with the COG complex. *Molecular Biology of the Cell*. 24:2907-2917.
- 6.Wakana Y, Villeneuve J, van Galen J, Cruz-Garcia D, <u>Tagaya M.</u>*Malhotra, V. (2013) Kinesin-5/Eg5 is important for transport of CARTS from the trans-Golgi network to the cell surface. *Journal of Cell Biology*. 202:241-250.



MEXT-Supported Program for the Strategic Research Foundation a Private Universities (2014-2018)

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases

Role of phospholipids and transport proteins in the organization and interplay of ER subdomains



Katsuko Tani, Ph.D.

Professor

Laboratory of Cell Signaling, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences. Tokyo University o Pharmacy and Life Sciences

Research summary

The endoplasmic reticulum (ER) is a multifunctional organelle with various subdomains. Proper localization of lipids and proteins is essential for the formation of these subdomains. In this project, I plan to analyze the localization, movement, and dynamics of phospholipids and proteins in each subdomain and disclose the interplay between the MAM and other ER subdomains.

- 1.Doi H, Ushiyama M, Baba T, <u>Tani K</u>, Shiina M, Ogata K, Miyatake S, Fukuda-Yuzawa Y, Tsuji S. Nakashima M, Tsurusaki Y, Miyake N, Saitsu H, Ikeda S, Tanaka F, Matsumoto N, *Yoshida K. (2014) Late-onset spastic ataxia phenotype in a patient with a homozygous DDHD2 mutation. *Scientific Reports*. 4:7132
- 2. Yonekawa S, Furuno A, Baba T, Fujiki Y, Ogasawara Y, Yamamoto A, Tagaya M, *Tani K. (2011) Sec16B is involved in the endoplasmic reticulum export of the peroxisomal membrane biogenesis factor peroxin 16 (Pex16) in mammalian cells. *Proc Natl Acad Sci U S A*. 108:12746-12751
- 3.*Tani K, Tagaya M, Yonekawa S, Baba T. (2011) Dual function of Sec16B: Endoplasmic reticulum-derived protein secretion and peroxisome biogenesis in mammalian cells. Cell Logist. 1:164-167
- 4.Arimitsu N, Kogure T, Baba T, Nakao K, Hamamoto H, Sekimizu K, Yamamoto A, Nakanishi H, Taguchi R, Tagaya M, *Tani K. (2011) p125/Sec23-interacting protein (Sec23ip) is required for spermiogenesis. FEBS Letter. 585:2171-21767
- 5. Linuma T, Aoki T, Arasaki K, Hirose H, Yamamoto A, Samata R, Hauri HP, Arimitsu N, Tagaya M, *Tani, K. (2009) Role of syntaxin 18 in the organization of endoplasmic reticulum subdomains. *Journal of Cell Science*. 122:1680-1690

A C T I V I T Y R E P O R T



MEXT-Supported Program for the Strategic Research Foundation a

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Shun Nagashima, Ph.D.

Assistant Professor

Analysis of MITOL-deficient mice

Laboratory of Molecular Biochemistry, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.

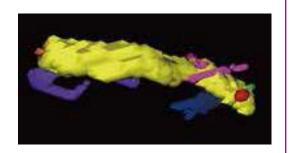
Research summary

Mitochondria are important organelle to produce energy, but mitochondrial dysfunction is closely involved in various diseases. There are several machineries regulating mitochondrial quality to maintain mitochondrial functions. Recently, it has been suggested that the efficient exchanges of lipids and calcium between mitochondria and ER by a direct tethering play an important role in maintaining the function of the mitochondria. We reported that mitochondrial ubiquitin ligase (MITOL) regulates mitochondria-ER contacts via Mfn2. In this project, we try to elucidate the mechanism of mitochondria-ER contact manner through analysis of MITOL-deficient mice.

Figure

Three-dimensional reconstruction of mitochondrion and ER.

Plural ER (red, blue, purple, pink, and green) contact with a mitochondrion (yellow).



- 1. Nagashima S., Yanagi S. (2014) Role of MITOL in mitochondrial dynamics and disease. Seikagaku. 86(1):63-7. Review. Japanese.
- 2. Nagashima S, Tokuyama T, Yonashiro R, Inatome R, Yanagi S. (2014) Roles of mitochondrial ubiquitin ligase MITOL/MARCH5 in mitochondrial dynamics and diseases. *J Biochem.* 155(5):273-9. Review
- 3.Sugiura, A., Nagashima, S. (equally contribution), Tokuyama, T., Amo, T., Matsuki, Y., Ishido, S., Kudo, Y., McBride, H.M., Fukuda, T., Matsushita, T., Inatome, R., and Yanagi, S. (2013) MITOL regulates endoplasmic reticulum-mitochondria contacts via Mitofusin2. *Mol. Cell* 51, 1-15.
- 4. Nagashima, S., Fukuda, T., Kubota, Y., Sugiura, A., Nakao, M., Inatome, R., and Yanagi, S. (2011) CRAG protects neuronal cells against cytotoxicity of expanded polyglutamine protein partially via c-fos-dependent AP-1 activation. *J. Biol. Chem.* 286(39), 33879-33889.
- 5.Sugiura, A., Yonashiro, R., Fukuda, T., Matsushita, N., Nagashima, S., Inatome, R., and Yanagi, S. (2011) A mitochondrial ubiquitin ligase MITOL controls cell toxicity of polyglutamine-expanded protein. *Mitochondrion* 11(1), 139-146

A C T I V I T Y R E P O R T



Role of mitochondria-associated membrane (MAM) in *Legionella* infection

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Kohei Arasaki, Ph.D.

Assistant Professor

Laboratory of Molecular Cell Biology, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences.

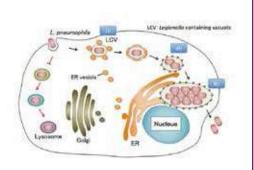
Research summary

Many microbes and viruses create intracellular environments advantageous for their survival and growth by hijacking host physiological machinery. Recent studies have revealed that some microbes and viruses manipulate function of organelle contact sites such as the ER (endoplasmic reticulum)-mitochondria contact site. In this project, I will try to understand the role of organelle contact sites, in particular, the ER-mitochondria contact site in intracellular pathogenesis of *Legionella pneumophila* that is known to cause severe pneumonia.

Figure

Intracellular pathogenesis of Legionella pneumophila.

After uptake into the host via phagocytosis, *Legionella pneumophila* prevents its degradation by inhibiting the delivery to lysosome. Simultaneously, Legionella recruits host ER-derived vesicles to the Legionella-containing vacuole to convert it into ER-Golgi intermediate compartment like structures, and then the pathogen-occupied membrane fuse with the ER and *Legionella* start to replicate.



- 1. Arasaki K, Shimizu H, Mogari H, Nishida N, Hirota N, Furuno A, Kudo A, Baba M, Baba N, Cheng J, Fujimoto T, Ishihara N, Ortiz-Sandoval C, Barlow L, Raturi A, Dohmae N, Wakana Y, Inoue H, Tani K, Dacks J, Simmen T, and Tagaya M. A novel role for the ancient SNARE Syntaxin 17 in regulating mitochondrial division. *Developmental Cell.* 2015 32(3): 304-17.
- 2.Hubber A, Arasaki K, Nakatsu F, Hardiman C, Lambright D, De Camilli P, Nagai H, Roy CR. The machinery at endoplasmic reticulum-plasma membrane contact sites contributes to spatial regulation of multiple Legionella effector proteins. *PLoS Pathog.* 2014 10(7): e1004222.
- 3. Arasaki K, Toomre DK, Roy CR. The Legionella pneumophila effector DrrA is sufficient to stimulate SNARE-dependent membrane fusion. *Cell Host Microbe*, 2012 11(1): 46-57.
- 4.Mukherjee S, Liu X, Arasaki K, McDonough J, Galán JE, Roy CR. Modulation of Rab GTPase function by a protein phosphocholine transferase. *Nature*. 2011 477(7362): 103-6.
- 5. Arasaki K. Roy CR. Legionella pneumophila promotes functional interactions between plasma membrane syntaxins and Sec22b. *Traffic.* 2010 11(5): 587-600.



Possible linkage between peroxisome biogenesis and lipid droplet formation through Sec16B

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Takashi Baba, Ph. D. Assistant Professor Laboratory of Cell Signaling, School of Life Sciences,

Research summary

The endoplasmic reticulum (ER) is the starting site of the secretory pathway. Recent studies revealed that the ER is adjacent to or in contact with other organelles such as mitochondria, peroxisomes, and lipid droplets. We recently reported that Sec16B, which was assumed to participate in the conventional secretory pathway, is involved in protein transport from the ER to peroxisomes. In this project, I will study a system for protein transport from the ER to peroxisomes and compare this with the conventional secretory system. I will also explore the relationship between peroxisome biogenesis and lipid droplet formation.

Tokyo University of Pharmacy and Life Sciences.

- 1. Baba T. Kashiwagi Y. Arimitsu N. Kogure T. Edo A. Maruyama T. Nakao K. Nakanishi H. Kinoshita M. Frohman MA, Yamamoto A. Tani K. (2014) Phosphatidic acid (PA) -preferring phospholipase A1 regulates mitochondrial dynamics. *J Biol Chem.* 289: 11497-11511.
- 2.<u>Baba T.</u>, Yamamoto A, Tagaya M, Tani K. (2013) A lysophospholipid acyltransferase antagonist, CI-976, creates novel membrane tubules marked by intracellular phospholipase A1 KIAA0725p. *Mol Cell Biochem.* 376: 151–161.
- 3.Inoue H*, <u>Baba T</u>*, Sato S, Ohtsuki R, Takemori A, Watanabe T, Tagaya M, Tani K. (2012) Roles of SAM and DDHD domains in mammalian intracellular phospholipase A1 KIAA0725p. *Biochim Biophys Acta*. 1823: 930-939. * equally contributed
- 4. Yonekawa S, Furuno A, <u>Baba T</u>, Fujiki Y, Ogasawara Y, Yamamoto A, Tagaya M, Tani K. (2011) Sec16B is involved in the endoplasmic reticulum export of the peroxisomal membrane biogenesis factor Peroxin 16 (Pex16) in mammalian cells. *Proc Natl Acad Sci U S A*. 108: 12746-12751.



Role of Phosphoinositide Turnover in MAM functions

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Kiyoko Fukami, Ph. D.

Professor

Laboratory of Genome and Biosignals,
Tokyo University of Pharmacy and Life Science.

Tokyo University of Pharmacy and

Research summary

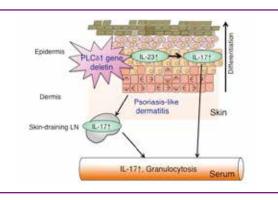
Phosphoinositide metabolism is an important intracellular signaling system involved in a variety of cell functions. In this system, phosphatidylinositol 4,5-bisphosphate is hydrolyzed by phospholipase C to generate two second messengers, inositol 1,4,5-trisphosphate (IP3) and diacylglycerol. IP3/calcium mobilization is suggested to be important in maintenance of the MAM function. On the other hand, dysfunction of MAM function often causes various diseases. Therefore we try to focus on the correlation between phosphoinositide metabolism and physiological functions of MAM. We also examine whether dysfunction of phosphoinositide metabolism in MAM links to cancer or skin diseases.

Figure

Importance of PLC δ 1 in skin.

Epidermal loss of PLC δ 1 causes increased production of IL-23 and IL-17 in the epidermis. This aberrant activation of the local IL-23/IL-17 axis resulted in a phenotype similar to that in human psoriasis. Serum IL-17 levels were also increased, resulting in granulocytosis.

(Kanemaru et al. Nature Commun. 2012)



- 1.Satow R., Hirano T., Batori R., Nakamura T., Murayama, Y. *Fukami K. (2014) Phospholipase C delta 1 induces E-cadherin expression and suppresses malignancy in colorectal cancer cells. *Proc. Natl. Acad. Sci. USA*. 111:13505-13510.
- 2.Nakamura Y., Kanemaru K., Kojima R., Hashimoto Y., Marunouchi T., Oka, N. Ogura T., Tanonaka K, *Fukami K. (2014) Simultaneous loss of phospholipase Cδ1 and phospholipase Cδ3 causes cardiomyocyte apoptosis and cardiomyopathy. Cell Death & Disease 5, e1215 doi:10.1038/cddis. 2014.181
 3.Hirano T., Satow R., Kato A., Tamura M., Murayama Y., Saya H., Kojima H., Nagano T., Okabe T., *Fukami K. (2013) Identification of novel small compounds that restore E-cadherin expression and inhibit tumor cell motility and invasiveness. Biochem. Pharmacol. 86:1419-29.
- 4.Kanemaru K., *Nakamura Y., Sato K., Kojima R., Takahashi S., Yamaguchi M., Ichinohe M., Kiyonari H., Shioi G., Kabashima K., Nakahigashi K., Asagiri M., Jamora C., Yamaguchi H., *Fukami K. (2012) Epidermal phospholipase Cδ1 regulates granulocyte counts and systemic interleukin-17 levels in mice. Nature Commun. 3:963. DOI:10.1038/ncomms1960.
- 5.*Yamaguchi H., Yoshida S., Muroi E., Yoshida N., Kawamura M., Kouchi Z., Nakamura Y., Sakai R., <u>Fukami K</u>. (2011) Phosphoinositide 3-kinase signaling mediated by p110α regulates invadopodia formation. *J. Cell Biol.* 193:1275-88.
- 6.Hirata M., Suzuki M., Ishii R., Satow R., Uchida T., Kitazumi T., Sasaki T., Kitamura T., Yamaguchi H., Nakamura Y., *Fukami K. (2011) Genetic defect in phospholipase Cδ1 protects mice from obesity by regulating thermogenesis and adipogenesis. *Diabetes* 60,:1926-37.



The roles of MAM in the activation of intestinal macrophages in inflammation

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Masato Tanaka, M.D., Ph. D.

Professor

Laboratory of Immune Regulation, School of Life Sciences, yo University of Pharmacy and Life Sciences.

Pharmacy and Life Sciences.

Research summary

Tissue-resident macrophages recognize dead cells or tissue damages in various physiological conditions, and initiate inflammation and immune responses. These responses by the macrophages are recently found to play important roles in determining the pathology of various diseases. We have currently identified the CD169-postive macrophages as the cells that orchestrate the immune responses associated with tissue damage. In this study, we are focusing on the roles of MAM in the activation of CD169-postive macrophages in inflammatory conditions, and are investigating the therapeutic strategy with regulating the function of MAM in these macrophages.

- 1.Karasawa K, Asano K, Moriyama S, Ushiki M, Monya M, Iida M, Kuboki E, Yagita H, Uchida K, Nitta K, <u>Tanaka M.</u> (2014) Vascular-resident CD169-positive Monocytes and Macrophages Control Neutrophil Accumulation in the Kidney with Ischemia-reperfusion Injury. *J. Am. Soc. Nephrol.* in press
- 2.Asano, K., Nabeyama, A., Miyake, Y., Qiu, CH., Kurita, A., Tomura, M., Kanagawa, O., Fujii, S., and <u>Tanaka, M.</u> (2011) CD169-positive macrophages dominate antitumor immunity by crosspresenting dead cell-associated antigens. *Immunity*, 34, 85-95.
- 3.Nabeyama, A., Kurita, A., Asano, K., Miyake, Y., Yasuda, T., Miura, I., Nishitai, G., Arakawa, S., Shimizu, S., Wakana, S., Yoshida, H., and <u>Tanaka, M.</u> (2010) xCT deficiency accelerates chemically induced tumorigenesis. *Proc. Natl. Acad. Sci. U S A*, 107, 6436-6441.
- 4.Miyake Y, Asano K, Kaise H, Uemura M, Nakayama M, <u>Tanaka M.</u> (2007) Critical role of macrophages in the marginal zone in the suppression of immune responses to apoptotic cell-associated antigens. *J Clin Invest* 117: 2268-2278.
- 5.Asano K, Miwa M, Miwa K, Hanayama R, Nagase H, Nagata S, <u>Tanaka M.</u> (2004) Masking of phosphatidylserine inhibits apoptotic cell engulfment and induces autoantibody production in mice. *J Exp Med*, 200: 459-467.



Roles of MAM during Endothelial-to-Mesenchymal Transition (EndMT) in Tumor Microenvironment

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Tetsuro Watabe, Ph. D.

Professor

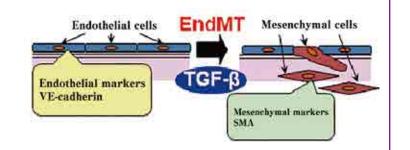
Laboratory of Oncology, School of Life Sciences, Tokyo University of Pharmacy and Life Science. Tokyo University of Pharmacy and

Research summary

Tumor tissues are composed not only of cancer cells but also of tumor vessels, cancer associated fibroblasts (CAFs) that play important roles in cancer progression. This tumor microenvironment is influenced by tumor specific cytokines that alter the structures of various organelle of tumor component cells. However, the roles of such tumor specific cytokines in the formation and maintenance of such organelle structures have not yet been elucidated. We attempt to study how transforming growth factor- β (TGF- β), which is abundant in tumor microenvironment, affect the mitochondria-associated membrane (MAM) of tumor endothelial cells. In tumor microenvironment, endothelial cells undergo endothelial-to-mesenchymal transition (EndMT), which leads to the formation of CAFs. This study will help understand the novel mechanisms how TGF- β -induced alteration of MAM is involved in the progression of cancer and aid developing new therapeutic strategies.

Figure

In tumor microenvironment, TGF- β induces endothelial-to-mesenchymal transition (EndMT) in which endothelial cells lose their characteristics (cell-cell contact and expression of endothelial markers, such as VE-cadherin) and acquire mesenchymal characteristics (high migratory activities and expression of mesenchymal markers such as smooth muscle α -actin).

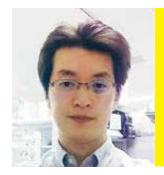


- 1.Miyazaki H, Yoshimatsu Y, Akatsu Y, Mishima K, Fukayama M, *Watabe T. Miyazono K. (2014) Expression of platelet-derived growth factor receptor β is maintained by Prox1 in lymphatic endothelial cells and is required for tumor lymphangiogenesis. Cancer Science. 2014 105:1116-1123
- 2.7. Yoshimatsu Y, Lee YG, Akatsu Y, Taguchi L, Suzuki HI, Cunha SI, Maruyama K, Suzuki Y, Yamazaki T, Katsura A, Oh SP, Zimmers TA, Lee SJ, Pietras K, Koh GY, *Miyazono K, Watabe T. (2013) Bone morphogenetic protein-9 inhibits lymphatic vessel formation via activin receptor-like kinase 1 during development and cancer progression. *Proc Natl Acad Sci U S A.* 110:18940-18945.
- 3.Kawata M, Koinuma D, Ogami T, Umezawa K, Iwata C, Watabe T, *Miyazono K. (2012) TGF-β-induced epithelial-mesenchymal transition of A549 lung adenocarcinoma cells is enhanced by pro-inflammatory cytokines derived from RAW 264.7 macrophage cells. Journal of Biochemistry. 151:205-216.
- 4.Mihira H, Suzuki HI, Akatsu Y, Yoshimatsu Y, Igarashi T, Miyazono K, *Watabe T. (2012) TGF-β-induced mesenchymal transition of MS-1 endothelial cells requires Smad-dependent cooperative activation of Rho signals and MRTF-A. *J Biochem.* 143:199-206.
- 5. Suzuki Y, Ohga N, Morishita Y, Hida K, Miyazono K, *Watabe T. (2010) BMP-9 induces proliferation of multiple types of endothelial cells in vitro and in vivo. *Journal of Cell Science*, 123:1684-1692.



In Vitro Reconstitution of the Mitochondria-Associated ER Membranes (MAM) using Xenopus egg extracts

Roles of Membrane Contact Sites in Organelle Dynamics and Diseases



Yoshitami Hashimoto, Ph. D.

Assistant Professor

Laboratory of Cellular Regulation,

Tokyo University of Pharmacy and Life Sciences.

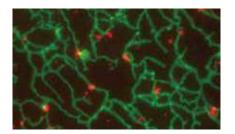
Tokyo University of Pharmacy and

Research summary

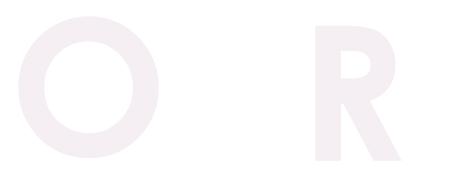
The endoplasmic reticulum (ER) is physically connected to mitochondria at a region called the mitochondria-associated membrane (MAM), which plays important roles in regulating not only mitochondrial activity but also various fundamental phenomena such as cell growth, apoptosis and autophagy. In this study, we will develop a novel in vitro system that recapitulate the MAM structure using fractionated *Xenopus* egg extracts for the purpose of analyzing the molecular architecture and biological function of the MAM.

Figure

A microscopic picture showing that mitochondria (red) are deposited along the ER tubule network (green) formed in egg extracts.



- 1.Ray Chaudhuri A, <u>Hashimoto Y</u>, Herrador R, Neelsen KJ, Fanchinetti D, Bermejo R, Cocito A, Costanzo V, Lopes M. (2012) Topoisomerase I poisoning results in PARP-mediated replication fork reversal. *Nat. Struct. Mol. Biol.*, 19, 417–23.
- 2. Hashimoto Y, Puddu F, Costanzo V. (2012) RAD51 and MRE11 dependent reassembly of uncoupled CMG helicase complex at collapsed replication forks. Nat. Struct. Mol. Biol., 19, 17–24.
- 3. Hashimoto Y, Ray Chaudhuri A, Lopes M, Costanzo V. (2010) Rad51 protects nascent DNA from Mre11-dependent degradation and promotes continuous DNA synthesis. *Nat. Struct. Mol. Biol.*, 17, 1305–1311.
- 4. Hashimoto Y, Tsujimura T, Sugino A, Takisawa H. (2006) The phosphorylated C-terminal domain of Xenopus Cut5 directly mediates ATR-dependent activation of Chk1. *Genes Cells*, 11, 993-1007.







Roles of Membrane Contact Sites in Organelle Dynamics and Diseases Newsletter Vol.1

Date of issue /April 2015

Domain Code : S1411014 (2014-2018) Organizer : Shigeru Yanagi

Tokyo University of Pharmacy and Life Science

Laboratory of Molecular Biocheistry 1432-1 Horinouchi, Hachioji-shi, Tokyo

192-0392, Japan

Issued by

TEL.042-676-7146 FAX.042-676-7149 E-mail: info@organella-tupls.net